

METABONOMICS

a generic platform for the investigation of drug toxicity, altered gene function and disease processes

Metabonomics can offer one of the most complete solutions to tackling one of the pharmaceutical industry's biggest problems – attrition. This article discusses the scope and uses of metabonomics in different areas of the drug discovery and development process.

Minimising 'attrition' is one of the most important aims of a drug discovery and development programme, and the application of novel technologies that increase the probability of making the right choice early saves resources, and promotes safety, efficacy and profitability in the pharmaceutical industry. Metabonomics¹⁻⁵ is a metabolic systems approach that is complementary to genomics and proteomics, which allows the characterisation of metabolic disease processes, genetic modifications and drug toxicity. In particular, metabonomics is focused on the study of altered whole organism metabolic control and homeostasis in complex systems where there are many interacting cell types performing different biochemical functions at the same time. Because of the integrative nature of the approach, metabonomics can add significant value to conventionally derived information on drug utility at several stages in the discovery and development process where complex system function needs to be understood^{1,2}. Studying the effects of drugs on whole organisms by metabonomics, relies

on multiparametric measurement of alterations in metabolism to a stressor or intervention. This and related metabolomic approaches can also be readily adapted to investigate the functional consequences of genetic modification⁶⁻⁸, which is potentially of great importance in the creation and validation of new models of human disease and efficacy. Thus, there is considerable scope for the application of metabonomic approaches within the pharmaceutical industry from discovery through clinical development and beyond specifically in the following areas:

Potential discovery applications

- Early *in vivo* toxicological testing and screening based on mechanisms.
- Lead compound selection and pre-lead prioritisation.
- *In vivo* efficacy screening in animal models.

Potential development applications

- Discovery of novel preclinical safety biomarkers and mechanisms.

By Professor Jeremy K. Nicholson

Figure 1

Relationships under study in the systems biology of complex organisms; only by integrating many levels of measurement can the relationships between gene complement and disease processes and their treatment be understood

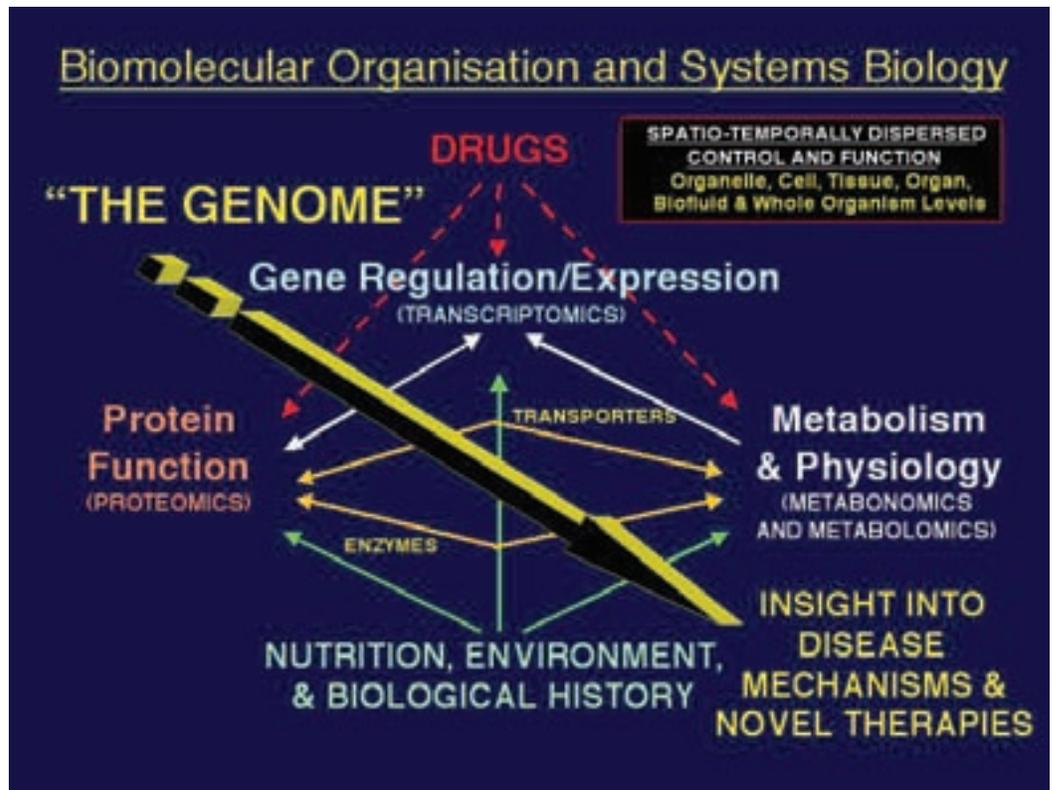
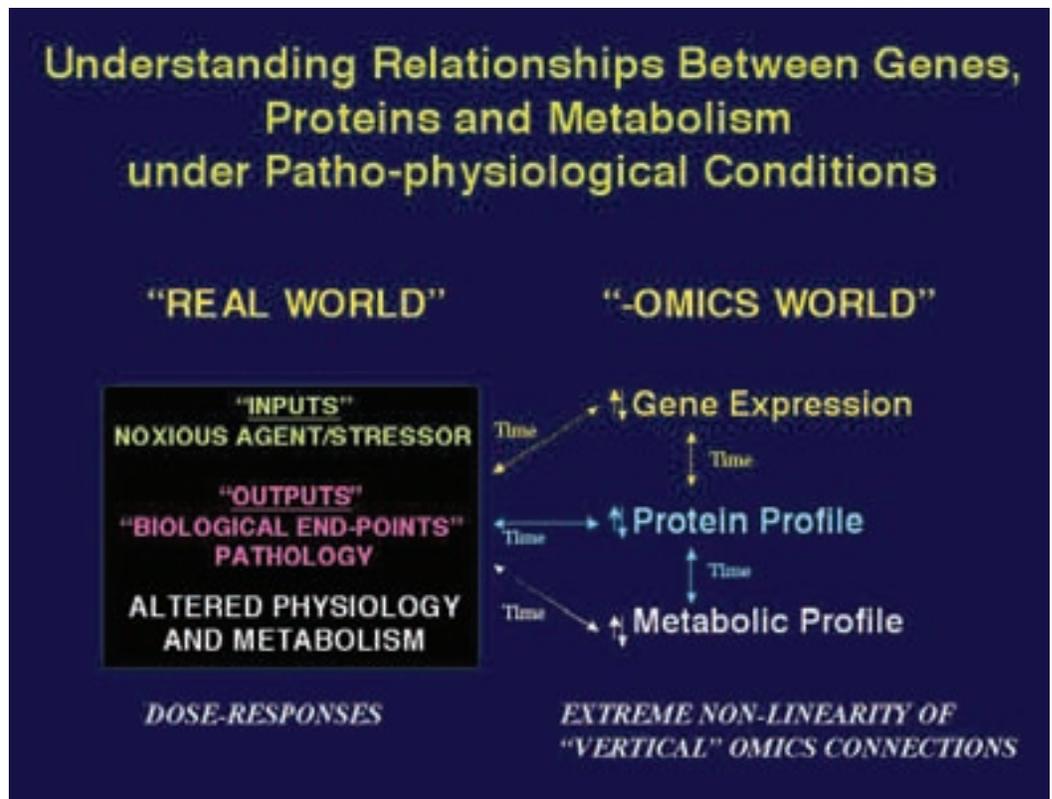


Figure 2

Relationships between real world inputs (stressors) and outputs and their time displaced connections with the measured “omics responses”. We seek to understand the connections between inputs and stressors and the outputs in terms of disease via making omics measurements, but modelling these interactions is complicated by the time and space displacement of the events. Also in the real world dose-response relationships are not mimicked by the non-linear relationships between variations at one omics level to another



- Metabolic validation of animal models against human disease profiles.
- Novel clinical safety and efficacy biomarker discovery.
- Pharmaco-metabonomic profiling to understand metabolic bases for adverse drug reactions.

The role of metabolic studies in understanding functional integrity of the whole organism

Knowing the genome sequence of humans or other species does not of itself explain the fundamental nature of most disease processes, and there is much interest in approaches that relate gene expression to phenotypic outcome. Several technologies are being developed to achieve this end, viz:

- Genetic complement and gene expression (genomics and transcriptomics).
- Protein synthesis and cell signalling (proteomics).
- Metabolic regulation of cellular systems and multivariate measurement and modelling of multivariate metabolic profiles (metabolomics).
- Systemic metabolic control and its disruption and physiological regulation in complex multicellular organisms (metabonomics).

In complex organisms, these features represent interdependent levels of biomolecular organisation and control which are also impacted by environmental events and stressors throughout life⁵. Some of the main relationships between the various levels of biomolecular organisation and the interactions between gene-proteins-metabolites and environmental factors that need to be studied to understand the whole organism and drug intervention are summarised in **Figure 1**. Note that there are many cell types interacting in mammals and their functional state is linked together by metabolic axes that communicate through the body fluids and that metabolic regulation of the whole system is dispersed in space and time. The characterisation of the different levels of system activity, by appropriate analytical methods, describes changes in biological activity as complex multivariate data sets that can be analysed using a variety of chemometric and bioinformatic tools³. The aim of such procedures is to extract latent biochemical information that is of diagnostic or prognostic value, and reflects actual biological events rather than potentials for disease offered by gene expression and proteomic data collected after exposure to a drug or stressor. It is thus necessary to relate real world or end-point observations to the measurements provided by the ‘-omics tech-

nologies’ (**Figure 2**) but modelling this is complicated by the time-displacement of many of the distributed interactions. This enables the understanding of the relationships between the inputs that change omics responses and the outputs of those responses. Metabolic changes are real-world end-points in their own right, whereas gene expression changes are not, merely giving the potential for an end-point change³. As such, metabonomics provides a useful practical connection between the other omics platforms and the real pathological end-points.

Metabonomics in drug toxicology

Metabonomics offers a complementary approach to genomics and proteomics that gives information on whole organism functional integrity over time following drug exposure⁵. Target tissues or processes and biomarkers can be identified via characteristic changes in the pattern of concentrations of endogenous metabolites in biofluids that relate to site and mechanism of toxicity.

A substantial body of work has been performed in this area using NMR spectroscopy as a lead tool. NMR is non-destructive of the sample and provides an unbiased overview of many important metabolites present in biosamples¹. ¹H NMR analysis of biofluids has uncovered novel metabolic markers of organ-specific toxicity in the laboratory rodent, and this ‘exploratory’ role is one in which biofluid NMR spectroscopy excels^{9,10}. However, it should be noted that other mass spectrometric-based techniques also provide powerful multivariate analytical probes especially when combined with Gas Chromatography⁸ or liquid chromatography. Indeed, the newly developed (by the Waters Corporation) approach of ultra performance (very high pressure liquid chromatography, UPLC™) offers many advantages in metabonomic analysis in terms of resolution and sensitivity, but is not yet fully commercially available.

Returning to NMR spectroscopy, which is currently the best-established method for pharmaceutical and clinical metabonomics^{1,3}, total potential biomarker information in NMR spectra of biofluids is very rich, as hundreds of compounds representing a variety of metabolic pathways can be measured using appropriate mathematical processing. The NMR spectrum of a biofluid can be conveniently divided into ‘biomarker windows’, which are spectral regions that often contain signals from metabolites associated with specific targets for toxicity, eg renal and hepatic⁹, as well as more subtle toxicological

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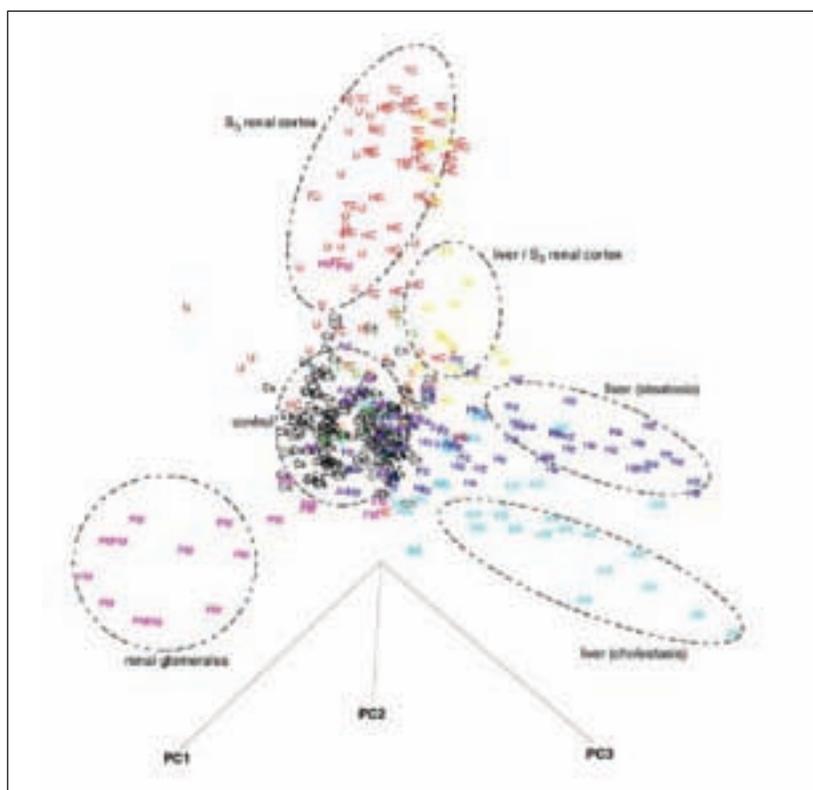


Figure 3

A simple principle components analysis of NMR data obtained from several toxin classes (each compound coded a different colour) in relation to the controls (black). Each coloured letter code point represents an individual animal mapped on to a simplified metabolic co-ordinate position (see references 1-5 for details). Distinct clustering is observed according to type of lesion which is a result of the perturbed NMR-detectable metabolic variation that is characteristic of the class

lesions such as phospholipidosis^{11,12}. Many such windows exist in NMR spectra and diagnostic information on hundreds of different types of disease or toxic processes (only a few of the known possibilities are shown) can potentially be derived from one measurement³.

While toxins may affect gene regulation or expression directly, significant responses may be completely unrelated to gene switching. In the latter case, genomic and proteomic methods are likely to be ineffective. However, all drug-induced effects involve disturbed endogenous metabolite concentration ratios, resulting from direct chemical reaction, altered binding to macromolecules, modified control mechanisms and induction or inhibition of enzymes. If these disturbances overwhelm compensatory or adaptive mechanisms, consequences recognised as toxicity occur. As metabolite concentrations in several key body fluids relate to cell and tissue processes, so are toxin- or disease-induced disequilibria reflected in those fluids. Using the pattern recognition approach, the NMR spectra can be used for:

- Classification of the sample as normal or abnormal (this is a useful tool in the control of spectrometer automation using sequential flow injection NMR spectroscopy).

- Classification of target organ toxicity and site of action within the organ. In certain cases, mechanism of toxic action may also be classified at this level.
- Identification of biomarkers of toxic effect of a compound.
- Evaluation of the time-course of effect, eg the evolution of the onset of toxicity and recovery.

The information derived from databases of NMR spectra can be maximised by use of appropriate chemometric and multivariate analytical strategies³. Preliminary analysis involves the application of unsupervised pattern recognition methods such as Principal Components Analysis (PCA) or cluster algorithms that assume no prior knowledge of sample class (Figure 3). Information relating to biomarkers of toxicity or recovery can be extracted from the analysis with a view to furthering the understanding of the mechanisms of toxicity and this has been well demonstrated with respect to failed drugs that have unusual mechanisms of toxicity¹³.

In August 2000 a major metabonomic toxicology project was commenced involving Imperial College and five drug companies (Pfizer, Hoffman La Roche, Lilly, Novo Nordisk and Bristol Myers Squibb): The Consortium on Metabonomic Toxicology¹⁴. The COMET project will not terminate until 2005 and by this time the time-related metabolic effects 147 compounds will have been studied in low and high dose conditions in rats and in some cases mice in order to generate novel metabolic biomarkers of drug toxicity and to enable the construction of expert systems for toxicological screening based on urine analysis. So far the results have been encouraging with the development of new methods for comparing toxicological similarities of unknowns to the large metabolic database and a high degree of success in determining the classification of toxicity of unknowns and test sets¹⁵. A major publication on the effectiveness of this approach as a screening tool is currently being prepared.

Metabonomics as a functional genomic tool

Metabonomics can be used to separate classes of experimental animals such as mice and rats according to their strain based on the endogenous metabolite patterns in their biofluids^{2,5,6}. This is due to the fact that differences in 'silent-gene' function between strains can influence the fluxes of metabolites through many key intermediary pathways resulting in distinct animal

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'metabotypes'⁶. Such differences may help explain differential toxicity of drugs between strains where the metabolic fate and hence potential toxicity of the drug itself is linked to the activity of endogenous pathways. There is also a strong indication for the use of metabonomics in the phenotyping of mutant or transgenic animals and the investigation of the consequences of transgenesis. In a recent study we have shown that the transfection process itself can cause significant metabolic differences in hepatoma cell lines due to cell membrane disruption in the host cell¹⁶. It is important to differentiate such 'unwitting' consequences of the genetic engineering process from the 'witting' or intended result, as these can potentially confuse the interpretation of the function of particular genes or gene classes when the cell system or organism is examined for the physical effects of the intervention. This is potentially important to pharmaceutical companies trying to genetically-engineer new animal models of disease using biochemically-invasive transfection procedures. Furthermore,

metabonomic approaches can give deep insight into the metabolic similarities or differences between mutant or K/O animals and the human disease processes that they are actually intended to simulate. If the veracity of an animal model can be established using metabolic criteria, ie biomarkers of the disease process, then it may also be possible to monitor the efficacy of novel therapeutic agents (normalisation of the biochemical profile) using metabonomic criteria. Such approaches may be of great future value to the pharmaceutical industry in the quest for the discovery of safe and efficacious new drugs.

Clinical metabonomics

In the clinic, metabonomics also offers promising new diagnostics and biomarker generation capabilities. For instance, it has been shown that unique combination markers of atherosclerosis can be discovered using combined spectroscopic and multivariate statistical analysis⁴ and that it is possible to determine the level of atherosclerotic damage (Figure 4) using minimally invasive blood

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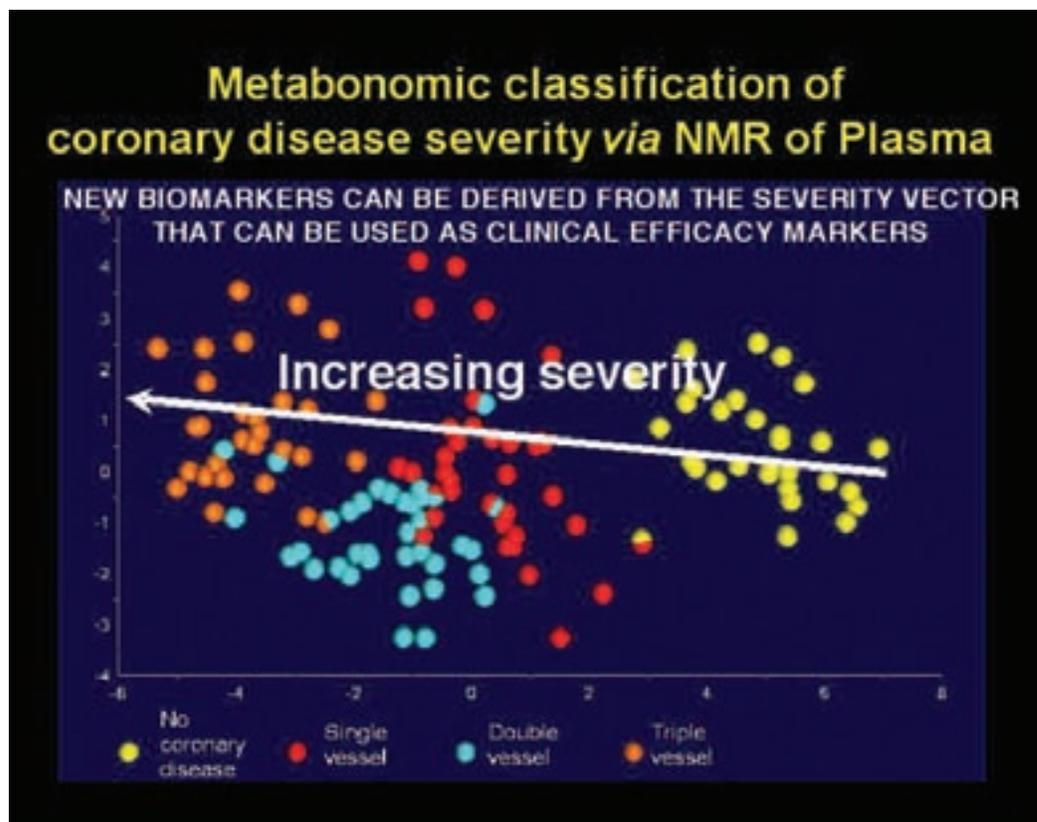
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Figure 4

A clinical metabonomic map of individual patients showing different levels of coronary artery disease (as determined by NMR spectroscopy of blood plasma and validated using angiographic criteria – colour-coded according to the number of obstructed coronary vessels determined for each patient)

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plasma analysis in patients as measured against 'gold standard' techniques such as angiography (itself a highly costly and potentially dangerous procedure). Such studies are not only valuable in the clinic as potentially important diagnostic aids, but also as a means for uncovering novel biomarkers that could be used for either quantitatively monitoring therapeutic efficacy of novel compounds. Also such models can generate new criteria with which to assess the metabolic similarities between human conditions and the animal models that are created to mimic them in pharmaceutical discovery and development studies. Overall, metabonomics offers one of the most complete solutions to tackling problems surrounding pharmaceutical attrition as it allows minimally invasive repeat studies in man and animals and the potential for translatability of data and models between species in a way not currently offered by genomics or proteomics. Furthermore, metabolic end-points are in the 'real world' and thus more readily related to clinical biochemical or pathological end-points than most other omics technologies. As such, metabonomics and related techniques are now being widely embraced by big pharma and the US FDA and EPA (both of whom have set up internal metabo-

nomics facilities in the last two years) as the next step forward in the battle of understanding complex system failure due to xenobiotic exposure or disease processes. **DDW**

Professor Jeremy Nicholson is Head of Biological Chemistry at Imperial College where he has pioneered the use of advanced spectroscopic and pattern recognition methods for metabolic profiling for more than 20 years. The author of more than 400 scientific papers, he leads an internationally renowned research team working on metabolic disease mechanisms. His work has been recognised by many awards including The Royal Society of Chemistry Silver Medal for Analytical Chemistry (1992) and Gold Medal for Analytical Science (1997), The Chromatographic Society Jubilee Silver Medal (1994), The Pfizer Academic Prize for Chemical and Medicinal Technologies (2002) and the Royal Society of Chemistry Medal for Chemical Biology (2003). Professor Nicholson is a Fellow of the Royal Society of Chemistry, The Institute of Biology and The Royal College of Pathologists.